

09PH
C174



Molecular and cellular studies examining the biological significance of different isoforms of the receptor tyrosine kinase, c-Kit

Antony Charles Cambareri
B. Sc. (Hons)

Institute of Medical and Veterinary Science

Department of Medicine
University of Adelaide
South Australia

A thesis submitted to the University of Adelaide
in candidature for the degree of Doctor of Philosophy
October 2004

Contents

1	INTRODUCTION	1
1.1.	C-KIT - A MEMBER OF THE GROWTH FACTOR RECEPTOR TYROSINE KINASE FAMILY	1
1.1.1.	<i>The Growth Factor Receptor Tyrosine Kinases.....</i>	1
1.1.2.	<i>Signalling through RTKs</i>	3
1.1.3.	<i>c-Kit in Cancer</i>	6
1.2.	HISTORY OF C-KIT.....	8
1.2.1.	<i>- The discovery of the c-Kit proto-oncogene.....</i>	8
1.2.2.	<i>Association of c-Kit with the W locus</i>	10
1.3.	THE LIGAND TO C-KIT: THE LINK BETWEEN W AND SL MUTANT MICE	13
1.4.	BIOCHEMICAL ANALYSIS OF C-KIT - LIGAND BINDING DOMAIN	16
1.5.	LIGAND INDUCED RECEPTOR DIMERISATION	18
1.6.	LIGAND INDUCED RECEPTOR SIGNALLING	20
1.6.1.	<i>SH2</i>	20
1.6.2.	<i>SH3</i>	21
1.6.3.	<i>PH.....</i>	22
1.6.4.	<i>PTB</i>	23
1.6.5.	<i>Modular Generation of Diversity.....</i>	23
1.7.	SIGNALLING THROUGH C-KIT	24
1.7.1.	<i>PI3-Kinase</i>	25
1.7.2.	<i>Phospholipase C-γ1</i>	26
1.7.3.	<i>RAS-MAPK</i>	27
1.7.4.	<i>Protein Kinase C.....</i>	28
1.7.5.	<i>Src Family Kinases</i>	29
1.7.6.	<i>Other</i>	29
1.8.	DOWMODULATION OF C-KIT EXPRESSION/ACTIVATION.....	30
1.8.1.	<i>PKC</i>	31
1.8.2.	<i>Extracellular domain shedding.....</i>	32
1.8.3.	<i>Ubiquitination.....</i>	32
1.8.4.	<i>Phosphatases</i>	33
1.8.5.	<i>Others</i>	34
1.9.	MECHANISMS INDUCING SIGNALLING DIVERSITY BY THE RTK	35
1.9.1.	<i>c-Kit Isoforms</i>	35
1.9.2.	<i>Heterodimerisation</i>	36
1.9.3.	<i>Receptor levels.....</i>	39
1.10.	AIMS.....	41
2	MATERIALS AND METHODS	43
2.1.	TISSUE CULTURE	43
2.1.1.	<i>Tissue Culture Media and Solutions</i>	43
2.1.2.	<i>Cytokines and growth factors</i>	46
2.2.	CULTURE MAINTENANCE OF CELLS	47
2.2.1.	<i>Psi 2 cell line maintenance</i>	47
2.2.2.	<i>FDC-P1 cell line maintenance</i>	48
2.2.3.	<i>NIH3T3 cell line maintenance</i>	48
2.2.4.	<i>Maintenance of Myb Immortalised Haemopoietic Cell (MIHC) lines</i>	48
2.2.5.	<i>Maintenance of MO7e</i>	49
2.2.6.	<i>Cryopreservation of Cells</i>	49
2.2.7.	<i>Thawing of Cryopreserved Cells</i>	50
2.2.8.	<i>Cytology, Cytochemistry and Histology.....</i>	50
2.2.9.	<i>Morphological characterisation of cells</i>	50
2.2.10.	<i>Phenotypic characterisation of cells – esterase staining.....</i>	51
2.3.	IMMUNOASSAYS	52
2.3.1.	<i>Antibodies</i>	52
2.3.2.	<i>Immunofluorescence Assay.....</i>	53
2.3.3.	<i>Alkaline Phosphatase Anti-Alkaline Phosphatase Technique.....</i>	54
2.3.4.	<i>Quantitative confocal microscopy for determination of endogenous mSCF levels.....</i>	56
2.3.5.	<i>Confocal Microscopy – visualisation of receptor internalisation.....</i>	57
2.4.	ANALYSIS OF CELL SURVIVAL, PROLIFERATION AND GROWTH BY PKH ASSAY	58

2.5.	SATURATION BINDING ANALYSIS (SCATCHARD ANALYSIS)	60
2.6.	PROTEIN ANALYSIS	61
2.6.1.	<i>Preparation of cellular lysates.....</i>	61
2.6.2.	<i>Immunoprecipitation.....</i>	62
2.6.3.	<i>Determination of the amount of protein within lysates.....</i>	62
2.6.4.	<i>Size determination of proteins – SDS PAGE</i>	63
2.6.5.	<i>Transfer of proteins to PVDF</i>	64
2.6.6.	<i>Visualisation of blots</i>	64
2.6.7.	<i>Quantitation of protein bands.....</i>	65
2.7.	MANIPULATION OF DNA.....	65
2.7.1.	<i>Restriction Endonuclease Digestion.....</i>	65
2.7.2.	<i>Electrophoresis of DNA</i>	65
2.7.3.	<i>Size determination and quantitation of DNA fragments</i>	66
2.7.4.	<i>Purification of DNA.....</i>	67
2.7.5.	<i>Dephosphorylation of DNA</i>	68
2.7.6.	<i>Ligation.....</i>	69
2.7.7.	<i>Production of chemical competent bacterial cells</i>	69
2.7.8.	<i>Transformation of chemical competent cells</i>	70
2.7.9.	<i>Expansion of plasmid DNA.....</i>	70
2.7.10.	<i>Polymerase Chain Reaction.....</i>	72
2.7.11.	<i>Generation of anti-sense murine SCF.....</i>	73
2.7.12.	<i>Chimaeric c-Kit Generation – Step 1 PCR conditions.....</i>	74
2.7.13.	<i>Chimaeric c-Kit Generation – Steps 2 and 3</i>	77
2.7.14.	<i>PCR Site Directed Mutagenesis of chimaeric c-Kit cDNA</i>	79
2.7.15.	<i>Q-PCR</i>	81
2.7.16.	<i>Sequencing of DNA.....</i>	83
2.7.17.	<i>Calcium phosphate transfection into NIH3T3 cells</i>	84
2.7.18.	<i>Retroviral infection of suspension cells by co-cultivation</i>	85
2.8.	MANIPULATION OF RNA	86
2.8.1.	<i>Total RNA Extraction.....</i>	86
2.8.2.	<i>Quantitation of RNA</i>	87
2.8.3.	<i>Random Oligonucleotide Priming – probe generation.....</i>	87
2.8.4.	<i>Northern Blot Transfer</i>	88
2.9.	REAGENTS	89
2.9.1.	<i>Immunocytochemistry, Immunohistochemistry and Immunofluorescence Reagents</i>	89
2.9.2.	<i>Reagents For Protein Analysis</i>	91
2.9.3.	<i>Molecular Biology Reagents.....</i>	93

3 TRANSFORMING POTENTIAL OF C-KIT - EFFECTS OF COPY NUMBER AND ISOFORM

3.1.	C-KIT RECEPTOR LEVELS	96
3.2.	ECTOPIC EXPRESSION OF C-KIT IN THE NIH3T3 MODEL: PREVIOUS FINDINGS IN THIS LABORATORY	97
3.2.1.	<i>Overexpression of c-Kit</i>	98
3.2.2.	<i>Transformation potential of Human c-Kit isoforms</i>	98
3.3.	ECTOPIC EXPRESSION OF MURINE C-KIT IN NIH3T3 MODEL	101
3.3.1.	<i>Quantitation of mu c-Kit surface expression</i>	101
3.3.2.	<i>Autocrine stimulation.....</i>	101
3.3.3.	<i>Human c-Kit Isoforms</i>	103
3.3.4.	<i>Establishment of c-Kit copy number on cell pools</i>	104
3.3.5.	<i>Affinity of isoforms of human c-Kit for SCF</i>	105
3.3.6.	<i>Transformation of NIH3T3 cells expressing c-Kit isoforms</i>	105
3.4.	BIOCHEMICAL ANALYSIS OF HUMAN C-KIT ISOFORMS SIGNALLING.....	106
3.4.1.	<i>Kinetics of activation of c-Kit isoforms, recruitment of p85 subunit of PI3-K</i>	106
3.4.2.	<i>Internalisation of c-Kit following SCF stimulation.....</i>	107
3.4.3.	<i>Downstream signalling from c-Kit isoforms</i>	108
3.5.	DISCUSSION	109
3.5.1.	<i>c-Kit receptor levels</i>	109
3.5.2.	<i>Hu-c-Kit isoforms</i>	112
3.5.3.	<i>Biochemical analysis</i>	113

3.5.4. Future experiments	117
4 DEVELOPMENT OF CHIMAERIC C-KIT ISOFORMS FOR EXPRESSION IN NEW MYB IMMORTALISED HAEMOPOIETIC CELL (MIHC) LINES.....	118
4.1. MYB-IMMORTALISATION OF HAEMOPOIETIC CELLS	119
4.1.1. <i>Myb gene family</i>	119
4.1.2. <i>c-Myb structure</i>	120
4.1.3. <i>Cellular targets of c-Myb</i>	123
4.1.4. <i>c-Myb regulation of proliferation and differentiation</i>	123
4.1.5. <i>Activated c-Myb immortalisation of primitive haemopoietic cells</i>	125
4.2. LYN – A MEMBER OF THE SRC FAMILY OF TYROSINE KINASES	125
4.2.1. <i>Src Family Kinase Structure</i>	126
4.3. LYN AND C-KIT	128
4.3.1. <i>Lyn knockout mice</i>	129
4.4. GENERATION OF HUMAN/MOUSE CHIMAERIC C-KIT.....	130
4.4.1. <i>Strategy</i>	131
4.4.2. <i>Cloning chi-c-Kit GNNK ± into pBS-SK</i>	133
4.4.3. <i>Mutation corrections</i>	134
4.4.4. <i>Generation of MIHC c57 and Lyn -/- (use of mIL-3)</i>	134
4.4.5. <i>Morphology of MIHC lines</i>	137
4.5. INTRODUCTION OF CHI C-KIT GNNK+/- INTO LYN -/- AND C57 MIHC	138
4.6. RESPONSE OF CHIMAERIC C-KIT TO HUMAN SCF IN C57 AND LYN -/- MIHCs	142
4.6.1. <i>Chimaeric c-Kit Internalisation in response to human SCF</i>	142
4.6.2. <i>Proliferation and Survival in response to Human SCF</i>	143
4.7. CHIMAERIC C-KIT DOWNSTREAM SIGNALLING ANALYSIS- LYN-/- VS C57 MIHCs	145
4.8. DISCUSSION	149
4.8.1. <i>MIHC Generation</i>	150
4.8.2. <i>Cellular responses to exogenous Human SCF</i>	152
4.8.3. <i>Further experiments</i>	154
5 ANALYSIS OF C-KIT ISOFORM EXPRESSION IN PRIMITIVE HUMAN HAEMOPOIETIC CELLS.....	156
5.1. EXPRESSION OF C-KIT ISOFORMS IN PRIMITIVE HAEMOPOIETIC CELLS.....	156
5.2. G-CSF MEDIATED STEM CELL MOBILISATION – INVOLVEMENT OF C-KIT	157
5.3. ANALYSIS OF GNNK+/- EXPRESSION IN NORMAL BONE MARROW MONONUCLEAR CELLS (MNCS).....	160
5.3.1. <i>Design and validation of “minor groove binder” (MGB)- TaqMan probes</i>	161
5.3.2. <i>Isolation of Normal Bone Marrow CD34+ subpopulations</i>	163
5.3.3. <i>QPCR analysis of Normal Bone Marrow subsets</i>	164
5.4. ANALYSIS OF GNNK+/- EXPRESSION IN MOBILISED CD34+ CELLS	166
5.4.1. <i>Isolation of Peripheral Blood CD34 positive stem cells</i>	166
5.4.2. <i>QPCR analysis of Peripheral Blood CD34 positive cells</i>	167
5.4.3. <i>Cell surface c-Kit levels in mobilised CD34 stem cells</i>	168
5.5. DISCUSSION	170
5.6. APPENDIX 1	174
5.6.1. <i>CD34</i>	174
5.6.2. <i>CD33</i>	176
5.6.3. <i>CD38</i>	177
5.6.4. <i>CD19</i>	178
5.6.5. <i>CD61</i>	179
5.6.6. <i>CD7</i>	180
5.6.7. <i>Glycophorin A</i>	181
6 GENERAL DISCUSSION	182
6.1. MURINE C-KIT RECEPTOR LEVELS	183
6.2. HU-C-KIT ISOFORMS.....	186
6.2.1. <i>Biochemical analysis</i>	186
6.3. DEVELOPMENT OF A NEW MODEL TO TEST C-KIT CELLULAR RESPONSES.....	190
6.3.1. <i>Myb Immortalised Haemopoietic Cell (MIHC) development</i>	190
6.3.2. <i>Cellular responses to exogenous Human SCF by chimaeric c-Kit in MIHC</i>	193

6.3.3. <i>Future directions for MIHC model</i>	196
6.4. ISOFORM EXPRESSION.....	196
6.5. CLOSING COMMENTS	199
7 BIBLIOGRAPHY.....	201

Abstract

c-Kit is a member of the Receptor Tyrosine Kinase Type III family and has four naturally occurring isoforms. The work presented in Chapter 3 utilised full-length human or murine c-Kit cDNA expressed in murine cells. Over-expression of normal c-Kit was capable of contributing to oncogenic transformation. The analysis of human c-Kit isoforms demonstrated dissociation of various indicators of transformation (anchorage independence, loss of contact inhibition, tumourigenicity) in the NIH3T3 cell model.

Biochemical analysis of the c-Kit signalling revealed qualitative and quantitative differences between the GNNK+ and GNNK- c-Kit isoforms. The GNNK- isoform was hyperphosphorylated more extensively and rapidly, and was also more efficiently ubiquitinated and degraded than the GNNK+ counterpart. PI3-K was recruited and activated equally by both isoforms. Phosphorylation of MAPK paralleled that of the c-Kit isoform's phosphorylation.

In Chapter 4, a new model was developed using a chimaeric human extracellular c-Kit/murine transmembrane + intracellular c-Kit. This new molecule, in conjunction with a murine Myb Immortalised Haemopoietic Cell (MIHC) line was used to investigate a number of biological outcomes stimulated by SCF simultaneously. A MIHC line lacking Lyn was also analysed.

Chimaeric c-Kit displayed the same signalling characteristics exhibited by its' full-length human counterpart. The model showed that the GNNK- isoform was superior

in its survival stimulus to GNNK+, but both were equivalent in promoting proliferation. The absence of Lyn reduced the ability of both isoforms to promote survival.

The aim of work in Chapter 5 was to elucidate the expression patterns of the c-Kit isoforms in subsets of normal human haemopoietic cells. Methodology was developed to detect GNNK+/- c-Kit mRNA from rare subsets of cells from bone marrow. As c-Kit is known to be down-modulated in mobilised peripheral blood stem cells, mobilised CD34+ cells were also investigated. In all haemopoietic cells analysed, there was no significant difference in expression patterns of the c-Kit isoforms, with all samples expressing approximately 90% of total c-Kit transcripts as the GNNK- isoform. c-Kit downmodulation observed in mobilisation of CD34+ cells was not influenced at the level of transcription, but at the protein level.