An analysis of DRONC function and its regulation of expression during *Drosophila* development

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A thesis submitted for the degree of Doctor of Philosophy, In the Faculty of Health Science, The University of Adelaide December 2004

Table of Contents

Abstract

Statement

Acknowledgments

Publications

Chapter 1: Introduction

1.1 Programmed cell death (PCD)1
1.2 Conservation of the cell death machinery
1.2.1 Caenorhabditis elegans 1
1.2.2 Mammals2
1.2.3 Drosophila melanogaster 5
1.2.3.1 A model system to study cell death5
1.2.3.2 The <i>Drosophila</i> PCD machinery7
1.3 <i>Drosophila</i> caspases
1.3.1 <i>DCP-1</i>
1.3.2 <i>DECAY</i> 10
1.3.3 <i>DAMM</i>
1.3.4 <i>DRICE</i> 11
1.3.5 <i>DREDD</i> 12
1.3.6 <i>STRICA</i>
1.3.7 <i>DRONC</i>
1.4 Regulation of DRONC activation15
1.5 Ecdysone-mediated PCD17

1.6 EcR/Usp binding elements and target genes 1	19
1.7 The ecdysone-induced genetic regulatory hierarchy2	21
1.8 The ecdysone-induced transcriptional hierarchy in the salivary glands2	1
1.9 Broad-Complex (BR-C)2	22
1.9.1 BR-C-mediated regulation of <i>dronc</i> expression	25
1.10 The role of E93 in PCD 2	25
1.10.1 Salivary gland histolysis and its regulation2	27
1.11 Caspase substrates and salivary gland destruction	31
1.12 The PCD genetic hierarchy and midgut cell death	32
1.13 Aims	34

Chapter 2: Materials and Methods

2	2.1 Drosophila stocks and crosses	
	2.1.1 Genetic Interaction crosses	35
	2.1.2 <i>dronc</i> promoter-reporter crosses	35
	2.1.3 <i>dronc</i> locus deletion complementation lines	. 36
2	2.2 Drosophila protocols	
	2.2.1 Staging of animals	.37
	2.2.2 Germline transformation	. 37
	2.2.3 Lethality tests	. 38
	2.2.4 Cell death detection	. 38
	2.2.5 Gamma Irradiation of Larvae	. 39
	2.2.6 Caspase cleavage assays	39
	2.2.7 Histology	. 39
	2.2.8 Detection of β -galactosidase expression	40

2.2.9 Quantitative β -galactosidase assays41
2.2.10 Drosophila cell culture 41
2.2.11 Preparation of nuclear extracts and EMSA41
2.2.12 Immunohistochemistry 42
2.2.13 Transmission electron microscopy (TEM)43
2.2.14 Haemocyte analysis 43
2.3 Drosophila cell line manipulations
2.3.1 Cryopreservation of <i>Drosophila</i> cell lines 44
2.3.2 Thawing cryopreserved <i>Drosophila</i> cells
2.3.3 <i>Drosophila</i> cell culture RNA interference (RNAi) 44
2.3.4 Transfection and luciferase assay45
2.4 Immunoblotting 46
2.5 RT-PCR
2.5 RT-PCR
 2.5 RT-PCR
 2.5 RT-PCR
2.5 RT-PCR.462.6 Standard DNA Manipulations462.6.1 Quantification of DNA.462.6.2 Separation of DNA fragments by electrophoresis.472.6.3 Restriction endonuclease digestion47
2.5 RT-PCR.462.6 Standard DNA Manipulations462.6.1 Quantification of DNA.462.6.2 Separation of DNA fragments by electrophoresis.472.6.3 Restriction endonuclease digestion472.6.4 Purification of DNA fragments47
2.5 RT-PCR.462.6 Standard DNA Manipulations462.6.1 Quantification of DNA.462.6.2 Separation of DNA fragments by electrophoresis.472.6.3 Restriction endonuclease digestion472.6.4 Purification of DNA fragments47a) Phenol/chloroform extraction.47
2.5 RT-PCR.462.6 Standard DNA Manipulations462.6.1 Quantification of DNA.462.6.2 Separation of DNA fragments by electrophoresis.472.6.3 Restriction endonuclease digestion472.6.4 Purification of DNA fragments47a) Phenol/chloroform extraction.47b) ULTRA-CLEAN [™] .48
2.5 RT-PCR.462.6 Standard DNA Manipulations462.6.1 Quantification of DNA.462.6.2 Separation of DNA fragments by electrophoresis.472.6.3 Restriction endonuclease digestion472.6.4 Purification of DNA fragments47a) Phenol/chloroform extraction.47b) ULTRA-CLEAN TM 482.6.5 End-filling DNA.49
2.5 RT-PCR. 46 2.6 Standard DNA Manipulations 46 2.6.1 Quantification of DNA. 46 2.6.2 Separation of DNA fragments by electrophoresis. 47 2.6.3 Restriction endonuclease digestion 47 2.6.4 Purification of DNA fragments 47 a) Phenol/chloroform extraction. 47 b) ULTRA-CLEAN [™] . 48 2.6.5 End-filling DNA. 49 2.6.6 Dephosphorylation of DNA. 49
2.5 RT-PCR
2.5 RT-PCR. 46 2.6 Standard DNA Manipulations 46 2.6.1 Quantification of DNA. 46 2.6.2 Separation of DNA fragments by electrophoresis. 47 2.6.3 Restriction endonuclease digestion 47 2.6.4 Purification of DNA fragments 47 a) Phenol/chloroform extraction 47 b) ULTRA-CLEAN [™] 48 2.6.5 End-filling DNA. 49 2.6.7 Ligation of DNA fragments 49 2.7 PCR amplification and sequencing of DNA

a) Taq polymerase amplification	
b) DyNAzyme™ amplification	50
c) High fidelity PCR	50
2.7.2 Mutagenesis PCR reactions	50
2.7.3 DNA sequencing	51
2.8 Harvesting of plasmid DNA from bacterial cultures	
2.8.1 Small-scale plasmid extraction	52
2.8.2 Large-scale plasmid extraction	52
2.9 RNA Analysis	
2.9.1 Quantification of total RNA preparations	53
2.9.2 RNA extraction	53
2.9.3 RNA gel electrophoresis	54
2.9.4 Northern blotting	55
a) RNA transfer	55
b) Probe radio-labelling	55
c) Hybridisation and signal detection	56
2.10 Transformation of chemically competent bacterial Cells	
2.10.1 Preparation of competent <i>E. coli</i> cells	
2.10.2 Transformation of chemically competent cells	57
2.11 Protein Analysis	
2.11.1 Determining protein concentrations	57
2.11.2 Protein extraction	57
2.11.3 Recombinant protein generation	58
2.11.4 <i>in vitro</i> translation	
2.11.5 SDS-PAGE and protein transfer	59

	2.11.6 Western blotting	.60
	2.11.7 Stripping Western blots	60
2	2.12 Plasmid constructs	
	2.12.1 Generation of <i>dronc</i> promoter-reporter <i>LacZ</i> constructs	. 60
	2.12.2 Additional constructs and vector	. 61
2	2.13 Primer list	
	2.13.1 <i>dronc</i> promoter-reporter construct primer list	. 62
	2.13.2 Complementation construct mutation primers	. 62
	2.13.3 Ecdysone Receptor mutation forward primer	63

Chapter 3: Drosophila Caspase DRONC is Required for Multiple Developmental Cell Death Pathways and Stress-Induced Apoptosis

3.1 Introduction
3.2 A hypomorphic <i>dronc</i> allele
3.3 <i>dronc</i> expression in <i>KGO2994</i> midguts66
3.4 <i>dronc</i> expression in <i>KGO2994</i> salivary glands68
3.5 Creation of a specific <i>dronc</i> mutant69
3.6 Generation of complementation transgenes70
3.7 $CG6685^4$ and $dronc^{d5}$ animals have different larval organ morphology
3.8 $CG6685^4$ and $dronc^{d5}$ animals die at different developmental stages
3.9 CG6685 ⁴ and dronc ^{d5} animals have different developmental delays and survival
rates to the late third instar stage
3.10 Cell death in larval structures is affected by loss of DRONC
3.11 DRONC is not required for larval midgut destruction
3.11.1 <i>dronc</i> mutant midgut destruction occurs at pupariation

3.11.2 <i>dronc</i> mutant midguts are TUNEL positive
3.11.3 Caspase activation is maintained in <i>dronc</i> ^{d5} midguts
3.11.4 Aspects of autophagy are absent in <i>dronc</i> ^{d5} midguts78
3.12 DRONC is required for salivary gland removal
3.12.1 <i>dronc</i> mutant animals have persistent salivary glands
3.12.2 <i>dronc</i> mutant salivary glands are deficient for TUNEL81
3.12.3 Effector caspase activation is DRONC-dependent in salivary glands 81
3.12.4 <i>dronc</i> mutant salivary gland cells have ultrastructural anomalies
3.12.5 The PCD genetic regulatory pathway is active in <i>dronc</i> ^{d5} salivary glands83
3.13 DRONC is required for RPR-, HID- and GRIM-induced cell death in the eye84
3.14 <i>dronc</i> is required for radiation-induced cell death
3.15 <i>dronc</i> ^{d5} animals have increased blood cells
3.16 Discussion

Chapter 4: Distinct promoter regions regulate spatial and temporal expression of the Drosophila caspase dronc

4.1 Introduction	89
4.2 <i>dronc</i> promoter-driven <i>LacZ</i> expression in embryos and adult ovaries	91
4.3 dronc regulation in midguts and salivary glands during metamorphosis	93
4.4 Expression of the promoter- <i>LacZ</i> transgenes in larval brain lobes	96
4.5 Temporal regulation of the <i>dronc</i> promoter	97
4.6 The Role of E74A, BR-C and E93 in the regulation of <i>dronc</i> expression	98
4.7 Discussion	02

Chapter 5: Identification of regulatory elements controlling the temporal and spatial expression of dronc 5.1 Introduction 106 5.2 Analysis between 2.8 and 1.1kb of dronc promoter 108 5.3 A region of the dronc promoter contains a putative midgut repressor 108 5.4 A region of the dronc promoter is required for the spatial regulation of dronc. 109 108 5.5 dronc promoter contains an EcR/Usp binding element (EcRBE) 110 5.6 An EcR/Usp binding site resides in the dronc promoter 111 5.7 The EcR-B1 isoform specifically binds the dronc promoter 113 5.9 dronc EcRBE is important for ecdysone-mediated dronc transcription 114 5.10 Discussion 115

Chapter 6: General Discussion	120
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Abstract

Correct development of multicellular organisms requires the programmed removal of supernumerary, redundant, or damaged cells, a process achieved by apoptosis. Apoptosis, or Programmed Cell Death (PCD) is executed by caspases, a highly conserved family of cysteine proteases. The removal of redundant larval tissues during metamorphosis is controlled by the steroid hormone ecdysone. Ecdysone signalling is mediated by the nuclear receptor heterodimer EcR/Usp, which in turn transcriptionally activates a host of transcription factors which then go on to regulate genes essential for PCD, like caspases. The apical caspase *dronc* is upregulated in the larval midgut and salivary glands prior to their destruction and is dependent on the BR-C and E93 transcription factors. To further understand the role of *dronc* in development, a *dronc* mutant fly was generated and a transgenic promoter-reporter strategy was employed to investigate *dronc* regulation.

Larval organs from *dronc* mutants lack dying cells and, when irradiated, fail to show a radiation-induced PCD response. The midguts from *dronc* mutants undergo apoptosis and have high caspase activity. These data indicate that a *dronc*-independent caspase activation pathway is active in the midgut. Salivary glands from *dronc* mutants failed to be removed and have reduced caspase activity. Consequently it is clear that the role of DRONC differs significantly between the midgut and salivary glands.

The employment of a transgenic *dronc* promoter-*LacZ* reporter system identified promoter regions essential for the correct temporal and spatial expression of *dronc*. A region of the *dronc* promoter between 1.1kb and 2.8kb has elements essential for *LacZ* expression in salivary glands. This region was also dependent on the BR-C and E93

viii

transcription factors for salivary gland expression. A functional ecdysone receptor binding element (EcRBE) was identified in the *dronc* proximal promoter. The EcR-B1 isoform directly binds this EcRBE and is necessary for correct *dronc* expression in the larval salivary glands.

This work revealed some novel findings regarding the role of DRONC in development and the availability of a specific *dronc* mutant now makes it possible to explore some of the recently published non-apoptotic roles of *dronc*. This work aids in understanding how nuclear hormones control transcription and shows *dronc* to be an ideal model gene to explore these molecular and genetic processes.

Statement

This thesis contains no material which has been accepted for any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge, contains no material previously published or written by any other person, except where due reference has been made. I give my consent for this thesis to be made available for loan and photocopying.

Acknowledgements

Firstly I must thank Professor Sharad Kumar who gave me the opportunity to work and study in his laboratory and under whose supervision I have endeavoured to complete my PhD studies. Without his example and efforts I would have neither published nor completed these studies. I am also very grateful for the persistence and support from all members of the lab, past and present, who helped me get through the tough times (Loretta, Stuart, Belinda, Linda and Andrew). Thanks to Dr Dimitrios Cakouros for practical guidance toward the direction and formulation of many parts of my research, and whom is responsible for all EMSA analyses and *I(2)mbn* cell line experiments in chapter 5. Also thankyou to Kathryn Mills who has participated in many aspects of the *dronc* mutant work, including the P-element excision, complementation construct cloning, acridine orange staining, genetic interaction and irradiation experiments.

Thanks must go to Dr Eric Baehrecke, Dr John Abrams and Dr Tetyana Shandala for scientific and methodological advice. Also I must acknowledge Drs Leonie Quinn and Helena Richardson without whose guidance and support I could not have progressed in science.

When in doubt I always depended on the rock solid support and encouragement of Maura and family. My beautiful daughters, Ashley and Harriet, came along to remind me that life is love and family and even when in despair I was still awash with good fortune.

xi

Publications

Daish, T. J., Mills, K., and Kumar, S. (2004). *Drosophila* Caspase DRONC Is Required for Specific Developmental Cell Death Pathways and Stress-Induced Apoptosis. Dev Cell *7*, 909-915.

Chew, S. K., Akdemir, F., Chen, P., Lu, W. J., Mills, K., **Daish, T.J.,** Kumar, S., Rodriguez, A., and Abrams, J. M. (2004). The Apical Caspase *dronc* Governs Programmed and Unprogrammed Cell Death in *Drosophila*. Dev Cell *7*, 897-907.

Cakouros, D., **Daish, T. J**., and Kumar, S. (2004a). Ecdysone receptor directly binds the promoter of the *Drosophila* caspase *dronc*, regulating its expression in specific tissues. J Cell Biol *165*, 631-640.

Cakouros, D., **Daish, T. J**., Mills, K., and Kumar, S. (2004b). An arginine-histone methyl transferase, CARMER, coordinates ecdysone-mediated apoptosis in *Drosophila* cells. J Biol Chem *279*, 18467-18471.

Quinn, L., Coombe, M., Mills, K., **Daish, T.J**., Colussi, P., Kumar, S., and Richardson, H. (2003). Buffy, a *Drosophila* Bcl-2 protein, has anti-apoptotic and cell cycle inhibitory functions. Embo J *22*, 3568-3579.

xii

Daish, T. J., Cakouros, D., and Kumar, S. (2003). Distinct promoter regions regulate spatial and temporal expression of the *Drosophila* caspase *dronc*. Cell Death Differ *10*, 1348-1356.

Cakouros, D., **Daish, T.J**., Martin, D., Baehrecke, E. H., and Kumar, S. (2002). Ecdysone-induced expression of the caspase DRONC during hormone-dependent programmed cell death in Drosophila is regulated by Broad-Complex. J Cell Biol *157*, 985-995.

Harvey, N. L., **Daish, T.J.**, Mills, K., Dorstyn, L., Quinn, L. M., Read, S. H., Richardson, H., and Kumar, S. (2001). Characterization of the *Drosophila* caspase, DAMM. J Biol Chem *276*, 25342-25350.