

Protein Tyrosine Phosphatase Pez:

Its role in the regulation of cell-cell adhesions

A thesis submitted in fulfilment of the requirement for the award of the degree

DOCTOR OF PHILOSOPHY

from

The University of Adelaide

by

Carol Wadham

Department of Medicine Faculty of Health Sciences March 2003

Abstract

The balance of tyrosine phosphorylation in the cell is maintained by the opposing actions of protein tyrosine kinases (PTKs) and protein tyrosine phosphatases (PTPs). Investigation into tyrosine phosphorylation was initially focused on the action of PTKs. However, research over the past decade has revealed that PTPs also play a key role in signal transduction.

The multi-protein complexes that constitute the cell-cell adhesions in endothelial and epithelial tissues are dynamically restructured in response to extracellular and intracellular signalling. Tyrosine phosphorylation is involved in the regulation of both adherens junctions and tight junctions. Inhibitors of PTPs have been shown to disrupt cell-cell adhesions indicating that PTPs are important in maintaining adhesion integrity.

The maintenance of a selectively permeable barrier is an essential function of endothelial cells, which are the cells that line the lumen of blood vessels. Therefore, it is important to understand the normal functioning of the proteins in the cell-cell adhesion complexes. The aims of this research were to ascertain which members of the PTP family are expressed in human umbilical vein endothelial cells (HUVEC) and to characterise a PTP that may potentially be involved in the regulation of cell-cell adhesions.

A homology screen identified a cytosolic phosphatase, PTP-Pez, to be highly expressed in HUVEC. The presence of the protein-protein interaction FERM domain (band 4.1, ezrin, radixin and moesin) at the N-terminus of Pez predicted its localisation to the plasma membrane. Specific antibodies showed that in confluent monolayers Pez is cytoplasmic and concentrated at intercellular junctions but the protein is nuclear in sub-confluent cells. The adherens junction protein β -catenin and the tight junction protein occludin were both identified as potential substrates of Pez using a "substrate-trapping" approach. Data showing that Pez bound to and dephosphorylated β -catenin *in vivo* further substantiated this. A truncated form of Pez lacking the catalytic domain acted as a dominant negative mutant inhibiting the dephosphorylation of its substrates at intercellular junctions and enhancing cell motility. Canine epithelial cells overexpressing Pez underwent an apparent epithelial to mesenchymal transition (EMT), a process typified by downregulation of cell-cell contacts. These findings indicate that Pez plays a role in the regulation of cell-cell adhesion.

TABLE OF CONTENTS

| THESIS DECLARATION | Ш |
|--|--|
| ACKNOWLEDGEMENTS | Ш |
| PUBLICATIONS ARISING FROM THIS THESIS | IV |
| ABSTRACT | v |
| LIST OF FIGURES | XI |
| LIST OF TABLES | XII |
| CHAPTER 1: INTRODUCTION | 2 |
| 1.1 Tyrosine phosphorylation | 2 |
| 1.2 Protein tyrosine kinases 1.2.1 Receptor tyrosine kinases 1.2.1.1 Phosphotyrosine binding proteins 1.2.2 Non-receptor tyrosine kinases 1.2.2.1 Non-receptor tyrosine kinase signalling pathways | 2 2 3 4 4 |
| 1.3 Protein tyrosine phosphatases 1.3.1 PTP superfamily 1.3.2 Catalytic mechanism | 5 5 6 |
| 1.4 The classical PTP family 1.4.1 Receptor-like PTPs 1.4.1.1 RPTP extracellular domains 1.4.2 Cytosolic PTPs 1.4.2.1 Non-catalytic domains of cytosolic PTPs | 7 10 10 11 11 |
| 1.5 PTP substrate specificity 1.5.1 Substrate Specificity of SHP-1 and SHP-2 1.5.2 Substrate specificity of PTP1B and TC-PTP | 12 12 13 |
| 1.6 PTP knockouts 1.6.1 SHP-1 deficient "motheaten" mice 1.6.2 CD45 deficient mice 1.6.3 PTP1B deficient mice 1.6.4 PTP LAR deficient mice 1.6.5 PTP delta deficient mice 1.6.6 PTPβ/ζ deficient mice 1.6.7 Analysis of knockout phenotypes | 14 14 15 15 16 16 16 16 17 |
| 1.7 Identification of PTP substrates 1.7.1 Two-hybrid screens 1.7.2 Substrate-trapping | 1 8 18 20 |
| 1.8 Regulation of PTPs 1.8.1 Transcriptional regulation | 21 21 |

| 1.8.2 Regulation by alternative-splicing1.8.3 Regulation by phosphorylation1.8.4 Regulation by subcellular localisation1.8.5 Regulation by oxidation | 21 22 24 25 |
|--|---|
| 1.9 Regulation of cellular functions by PTPs 1.9.1 PTP regulation of cytokine signalling 1.9.2 PTP regulation of cell adhesion 1.9.2.1 Cell-matrix adhesion 1.9.2.2 Cell-cell adhesion 1.9.2.2.1 Adherens Junctions 1.9.2.2.1.1 Strength of adhesion | 26 26 29 30 30 32 |
| 1.9.2.2.1.2 Tyrosine phosphorylation and cell-cell adhesion 1.9.2.2.1.3 PTPs and adherens junctions 1.9.2.2.2 Tight Junctions 1.9.2.2.2.1 Assembly of the tight junction complex 1.9.2.2.2 Tyrosine phosphorylation and tight junction regulation | 33 33 34 35 36 |
| 1.10 Tyrosine phosphorylation and human disease 1.10.1 Adherens junctions and cancer 1.10.2 Tyrosine phosphorylation of adherens junction proteins and cancer 1.10.3 PTPs and cancer 1.10.4 PTPs and diabetes 1.10.5 PTPs and other diseases | 37 37 38 38 39 42 |
| 1.11 Concluding remarks and objectives | 42 |
| CHAPTER 2: MATERIALS AND METHODS | 46 |
| 2.1 Reagents 2.1.1 Antibodies 2.1.2 Solutions and buffers 2.1.3 Tissue culture reagents 2.1.4 Transfection of cells | 46 46 46 48 48 |
| 2.2 Tissue Culture 2.2.1 Cell Lines 2.2.2 Cryopreservation of cells 2.2.3 Isolation and culture of human umbilical vein endothelial cells 2.2.4 Culturing stable cell lines | 49 49 49 50 50 |
| 2.3 RNA isolation 2.3.1 Cell lysis 2.3.2 Phase separation 2.3.3 RNA precipitation | 52 52 52 52 |
| 2.4 Northern blotting 2.4.1Formaldehyde agarose gel for RNA 2.4.2 Sample preparation 2.4.3 Gel staining 2.4.4 Radioactive labelling of cDNA probes 2.4.5 Hybridisation 2.4.5.1 SSC hybridisation method 2.4.5.2 ExpressHyb[™] method | 53 53 53 53 54 54 54 54 55 |
| 2.5 cDNA synthesis and sequencing by reverse transcription and PCR amplification 2.5.1 First strand cDNA synthesis 2.5.2 PCR reaction 2.5.3 Specific amplification of PTP sequences from first strand cDNA | 55 55 56 57 |

| | 2.5.4 Ligation of PCR product into pGEM –T easy vector (Promega) 2.5.5 Transformation of competent JM109 | 57 57 |
|---|---|-----------------|
| | 2.6 cDNA probes | 58 |
| | 2.7 Pez DNA constructs | 58 |
| ÷ | 2.8 SDS-PAGE and Western blotting | 62 |
| | 2.8.1 Cell lysis | 62 |
| | 2.8.2 Subcellular fractionation | 63 |
| | 2.8.3 Determination of protein concentration of cell lysates | 63 |
| | 2.8.4 SDS-PAGE | 63 |
| | 2.8.5 Western blotting | 64 |
| | 2.9 Tyrosine-phosphatase assay | 66 |
| | 2.10 Enriching for tyrosine phosphorylated proteins | 67 |
| | 2.11 Substrate-trapping in vitro | 68 |
| | 2.11.1 PTP-Pez-coupled protein A-sepharose | 68 |
| | 2.11.2 Substrate-trapping | 68 |
| | 2.11.3. Scale Up for Identification of Substrate by Mass-Spectrometry | 68 |
| | 2.12 Co-immunoprecipitations | 60 |
| | 2.12.1 Co-immunoprecipitation with Pez | 69 69 |
| | 2.13 In vivo tyrosine phosphorylation in transiently transfected A431 cells | 69 |
| | | |
| | 2.14 In vivo tyrosine phosphorylation | 70 |
| | 2.14.1 In vivo tyrosine phosphorylation in stable MDCK cell lines | 70 |
| | 2.14.2 In vivo tyrosine phosphorylation in stable HEK293 cell lines | 70 |
| | 2.15 Microscopy | 70 |
| | 2.15.1 Confocal microscopy | 70 |
| | 2.15.2 Immunofluorescence | 71 |
| | 2.16 Permeability assays | 72 |
| | 2.17 Luciferase reporter assays | 72 |
| | 2.18 Wounding assay | 73 |
| | CHAPTER 3: RT-PCR ANALYSIS OF PTPS EXPRESSED IN AN ENDOTHELIAL | |
| | CELL LINE | 75 |
| | 3.1 Introduction | 75 |
| | 3.1.1 Protein tyrosine phosphatases in endothelial cells | 76 |
| | 3.1.1.1 Approach: | 78 |
| | 3.1.1.2. Criteria used for the selection of PTPs for further characterisation: | 78 |
| | 3.2 Results | 79 |
| | 3.2.1 PTP homology screen of HUVEC mRNA | 79 |
| | 3.2.2 Analysis of the PTP clones | 79 |
| | 3.2.3 Selecting a PTP to characterise | 80 |
| | 3.2.3.1 PTP-Pez | 82 |
| | 3.2.4 Confirmation of high PTP-Pez expression in HUVEC | 86 |
| | 3.2.4.1 Selection of sequence as a probe for Northern analysis | 86 |
| | 3.2.4.2 Analysis of regulation of PTP mRNA expression by growth factors and cytokines | 88 |
| | giornal and of the second of the second of growing and of toxinos | 00 |

CHAPTER 4: SUBCELLULAR LOCALISATION OF PTP-PEZ

| 4.1 Subcellular localisation of Pez | 92 |
|---|-----|
| 4.2 Methods | 93 |
| 4.2.1 Generation of antibodies | 93 |
| 4.2.1.1 Coupling of peptides | 93 |
| 4.2.1.2 Immunisation of rabbits | 93 |
| 4.2.1.3 Assay of the test-bleeds by ELISA | 94 |
| 4.2.2 Affinity purification of antibodies | 95 |
| 4.2.2.1 Coupling peptides to BSA | 95 |
| 4.2.2.2 Dialysis | 95 |
| 4.2.2.3 Coupling peptides-BSA to affigel | 96 |
| 4.3 Results | 97 |
| 4.3.1 Generation of antibodies against PTP-Pez peptides | 97 |
| 4.3.2 Characterisation of the antisera against Pez | 97 |
| 4.3.2.1 The antisera bind specifically to the peptides used for immunisation | 97 |
| 4.3.2.2 The antisera bind to the whole Pez protein. | 99 |
| 4.3.3 Determination of the subcellular localisation of Pez | 102 |
| 4.3.3.1 Pez subcellular localisation is cell-density dependent | 102 |
| 4.3.3.2 Nuclear localisation of Pez is serum dependent | 108 |
| 4.3.3.3 Pez translocates into the nucleus in cells at a 'wound edge': Inhibition by TGF β . | 110 |
| 4.3.4 Mechanism of nuclear localisation of Pez | 112 |
| 4.3.4.1 Background | 112 |
| 4.3.4.2 Deletion of the FERM domain reduces nuclear accumulation of Ectopic Pez | 113 |
| 4.3.4.3 Serine threonine phosphorylation regulates Pez translocation | 115 |
| 4.4 Conclusions | 117 |

CHAPTER 5: IDENTIFICATION OF POTENTIAL SUBSTRATES OF PTP-PEZ USING A SUBSTRATE-TRAPPING STRATEGY 123

| 5.1 Introduction | 123 |
|---|-----|
| 5.2 Results | 125 |
| 5.2.1 PTP-Pez phosphatase activity | 125 |
| 5.2.2 Trapping substrates of PTP-Pez | 129 |
| 5.2.3 Identification of substrates | 130 |
| 5.3 Discussion | 134 |
| CHAPTER 6: PTP-PEZ LOCALISES WITH THE ADHERENS JUNCTION | |
| COMPLEXES | 137 |
| 6.1 Introduction | 137 |
| 6.1.1 Cell-cell adhesion | 137 |
| 6.1.2 Tyrosine Phosphorylation and the Cell-Cell Adhesion Complexes | 139 |
| 6.1.3 Minimum essential criteria to establish that β -catenin is a Pez substrate | 139 |
| 6.2 Results | 140 |
| 6.2.1 PTP-Pez Interacts With β-catenin In Vivo | 140 |
| 6.2.1.1 Co-immunoprecipitation of endogenous Pez with β -catenin | 140 |
| or and the second | |

6.2.1.2 Truncation mutants of Pez co-immunoprecipitate with β -catenin

6.2.1.2 wt-Pez and deletion mutants of Pez localise to the cell-cell junctions in MDCK cells

142

144

| 6.2.2 Functions of Pez elucidated by ectopic expression of dominant negative mutants 6.2.2.1 ΔPTP-Pez acts as a dominant negative mutant and results in increased tyrosine phosphorylation. 6.2.2.2 Western analysis shows an increase in tyrosine phosphorylation of specific proteins by ΔPTP-Pez MDCK cells 6.2.2.3 Expression of ΔPTP-Pez in A431 cells | 147 147 2 in 149 151 |
|--|----------------------------------|
| 6.2.3 Overexpression of the dominant negative mutant (ΔPTP -Pez) enhances cell migration | 153 |
| 6.3 Discussion | 153 |
| CHAPTER SEVEN: PTP-PEZ AND THE TIGHT JUNCTION COMPLEXES | 161 |
| 7.1 Introduction | 161 |
| 7.1.2 Composition of the tight junctions | 161 |
| 7.1.3 The MAGUK family | 162 |
| 7.1.4 Occludin | 163 |
| 7.1.5 Claudins | 164 |
| 7.1.6 Junctional adhesion molecule (JAM) proteins | 165 |
| 7.1.7 Other tight junction proteins | 166 |
| 7.1.8 Tyrosine phosphorylation and tight junction permeability | 166 |
| 7.2 Results | 167 |
| 7.2.1 Association of Pez and occludin in vivo | 167 |
| 7.2.2 ZO-1 Co-immunoprecipitates with Pez | 168 |
| 7.3 Discussion | 168 |
| CHAPTER 8: OVEREXPRESSION OF PTP-PEZ INDUCES AN APPARENT | |
| | 175 |
| 8.1: Introduction | 175 |
| 8.2 Results | 176 |
| 8.2.1 Pez induces an EMT in MDCK cells | 176 |
| 8.2.2 Expression of Pez in MDCK cells | 180 |
| 8.2.3 Downregulation of cell-cell adhesion proteins following EMT | 180 |
| 8.2.4 No evidence of the activation of β -catenin/TCF/LEF signalling by Pez expression | 182 |
| 8.3 Discussion | 185 |
| 8.3.1 Background | 185 |
| 8.3.1.1 Epithelial to Mesenchymal Transition | 185 |
| 8.3.1.2 Activators of EMT | 185 |
| 8.3.1.3 Loss of cell-cell adhesion | 186 |
| 8.3.1.4 Transcriptional control of EMT | 187 |
| 8.3.1.4.1 The Slug/snail transcription factor family | 187 |
| 8.3.1.4.2 The β-catenin/TCF/LEF pathway. | 188 |
| 8.3.2 Summary and conclusions from this study | 189 |
| CHAPTER 9:GENERAL CONCLUSIONS AND DISCUSSION | 193 |
| 9.1 Conclusions | 193 |
| 9.2 Future Directions | 201 |
| | 8 85.2- |
| REFERENCES | 205 |