PUBLISHED VERSION

Greg R. Guerin, Ed Biffin, Zdravko Baruch, Andrew J. Lowe **Identifying centres of plant biodiversity in South Australia** PLoS One, 2016; 11(1):e0144779-1-e0144779-19

© 2016 Guerin et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Originally published at: http://doi.org/10.1371/journal.pone.0144779

PERMISSIONS				
http://creativecommons.org/licenses/by/4.0/				
© creative commons				
Attribution 4.0 International (CC BY 4.0)				
This is a human-readable summary of (and not a substitute for) the <u>license</u> . Disclaimer				
You are free to:				
Share — copy and redistribute the material in any medium or format Adapt — remix, transform, and build upon the material				
for any purpose, even commercially.				
The licensor cannot revoke these freedoms as long as you follow the license terms.				
Under the following terms:				
Attribution — You must give <u>appropriate credit</u> , provide a link to the license, and <u>indicate if changes were made</u> . You may do so in any reasonable manner, but not in any way that suggests the licensor endorses you or your use.				
No additional restrictions — You may not apply legal terms or <u>technological measures</u> that legally restrict others from doing anything the license permits.				



Citation: Guerin GR, Biffin E, Baruch Z, Lowe AJ (2016) Identifying Centres of Plant Biodiversity in South Australia. PLoS ONE 11(1): e0144779. doi:10.1371/journal.pone.0144779

Editor: RunGuo Zang, Chinese Academy of Forestry, CHINA

Received: October 7, 2015

Accepted: November 23, 2015

Published: January 6, 2016

Copyright: © 2016 Guerin et al. This is an open access article distributed under the terms of the <u>Creative Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: The plot-based species inventory datasets used in this study are available for download at <u>http://portal.aekos.org.au/dataset/</u> <u>168004</u> (Department of Environment, Water and Natural Resources 2014) and <u>http://portal.aekos.org.</u> <u>au/dataset/172373</u>. Herbarium records used in this study are publicly available from Australia's Virtual Herbarium (AVH: <u>http://avh.chah.org.au</u>). R scripts for georeferenced weighted endemism and georeferenced phylogenetic endemism functions are available at github.com/GregGuerin. Map layers showing remnant vegetation as polygons used in this study are available via Mapland (Department of Environment, Water and Natural Resources; see **RESEARCH ARTICLE**

Identifying Centres of Plant Biodiversity in South Australia

Greg R. Guerin¹*, Ed Biffin^{1,2}, Zdravko Baruch¹, Andrew J. Lowe^{1,2}

1 Terrestrial Ecosystem Research Network, Adelaide-node, School of Biological Sciences, University of Adelaide, North Terrace, Adelaide, South Australia 5005, Australia, 2 Science, Monitoring and Knowledge, Department of Environment, Water and Natural Resources, Adelaide, South Australia, Australia

* greg.guerin@adelaide.edu.au

Abstract

We aimed to identify regional centres of plant biodiversity in South Australia, a sub-continental land area of 983,482 km², by mapping a suite of metrics. Broad-brush conservation issues associated with the centres were mapped, specifically climate sensitivity, exposure to habitat fragmentation, introduced species and altered fire regimes. We compiled 727,417 plant species records from plot-based field surveys and herbarium records and mapped the following: species richness (all species; South Australian endemics; conservation-dependent species; introduced species); georeferenced weighted endemism, phylogenetic diversity, georeferenced phylogenetic endemism; and measures of beta diversity at local and state-wide scales. Associated conservation issues mapped were: climate sensitivity measured via ordination and non-linear modelling; habitat fragmentation represented by the proportion of remnant vegetation within a moving window; fire prone landscapes assessed using fire history records; invasive species assessed through diversity metrics, species distribution and literature. Compared to plots, herbarium data had higher spatial and taxonomic coverage but records were more biased towards major transport corridors. Beta diversity was influenced by sampling intensity and scale of comparison. We identified six centres of high plant biodiversity for South Australia: Western Kangaroo Island; Southern Mount Lofty Ranges; Anangu Pitjantjatjara Yankunytjatjara lands; Southern Flinders Ranges; Southern Eyre Peninsula; Lower South East. Species composition in the arid-mediterranean ecotone was the most climate sensitive. Fragmentation mapping highlighted the dichotomy between extensive land-use and high remnancy in the north and intensive land-use and low remnancy in the south. Invasive species were most species rich in agricultural areas close to population centres. Fire mapping revealed large variation in frequency across the state. Biodiversity scores were not always congruent between metrics or datasets, notably for categorical endemism to South Australia versus georeferenced weighted endemism, justifying diverse approaches and cautious interpretation. The study could be extended to high resolution assessments of biodiversity centres and cost:benefit analysis for interventions.



http://www.environment.sa.gov.au/Science/mapland). Interpolated climate layers used in this study are freely available via WorldClim (<u>http://www.worldclim.</u> org).

Funding: This research was supported by the Terrestrial Ecosystem Research Network and the South Australian Department of Environment, Water and Natural Resources. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Introduction

Biodiversity conservation requires application of limited resources, and spatial management priorities are generally based on values, threats and benefits at the level of ecological communities rather than individual species [1]. Identifying areas with high biodiversity value is therefore a fundamental layer of conservation planning, upon which landscape management and its relative costs and benefits can be super-imposed [2,3]. Biodiverse areas are defined on the basis of biological richness or uniqueness, which could relate to the presence of threatened, range-restricted or complementary species and ecological communities [2].

In the Mediterranean Biome, biodiverse landscapes face complex stressors [4] and basic conservation planning requires information on biodiversity values but also, for example, sensitivity and exposure to changes in climatic, land-use, invasion and fire regimes and the cost for return of management actions [1,5]. In South Australia, vegetation remnancy in some regions is low [6] and the climate is predicted to become significantly warmer and drier this century [in the order of +2°C and -10% MAP relative to a 1986–2005 baseline for intermediate emissions pathways; [7]). Although combining information on spatial biodiversity, climate sensitivity and habitat fragmentation can inform conservation priorities [8], it is rarely attempted at macro-ecological scales.

Here, we map coarse resolution plant biodiversity within the context of the state of South Australia using objective, landscape-scale metrics and identify areas of exceptional biodiversity and their associated conservation issues for context. Future extensions to this analysis could include higher resolution analysis over selected areas or in-depth analysis of ecosystem condition, threats and management costs.

Geopolitical context

This study is sub-continental and bound within South Australia, occupying the central, southern Australian mainland and off-shore islands (Fig 1). From an ecological perspective, this area is arbitrary, as a number of the state's bioregions (defined by 'IBRA', the Interim Biogeographic Regionalisation for Australia, [9]; Fig A in <u>S1 Appendix</u>) overlap state borders. However, there are two reasons for political bounds: **1**. The existence of a large, yet to date largely untapped, vegetation inventory dataset, the *Biological Survey of South Australia*, which was deliberately implemented to sample the state's ecological diversity and includes systematically sampled presence/absence data [10]. **2**. Ecosystems are primarily managed at state level in Australia and there is a practical need to provide information for biodiversity conservation in this context.

Biodiversity assessments for South Australia

Several studies have mapped biodiversity metrics for specific taxa across the Australian continent (e.g. Anura (frogs) [11]; *Acacia* [12]; bryophytes [13]; *Acacia* [14]; *Glycine* [15]; *Eucalyptus* [16], generally revealing idiosyncratic patterns among taxa for South Australia. Crisp *et al.* [17] and Laffan & Crisp [18] used 118,643 Australia-wide herbarium records of 8,560 vascular plants species to identify centres of endemism at a scale of 1° grid cells. For South Australia, they identified the Adelaide–Kangaroo Island area as having high endemism at continental scale. An assessment of the evolutionary uniqueness of the Australian arid biota revealed several South Australian areas as being continental refugia, including the Flinders Ranges and Kangaroo Island [19]. Traill *et al.*, [20] analysed data from relevés in the Alinytjara Wilurara Natural Resource Management (AW NRM) region, a desert area in the north-west of South Australia. By partitioning species diversity among *a priori* "major vegetation groups" based on representative plots, they concluded that climate averaged within vegetation groups was not a good predictor. State and Federal Governments have also assessed 'hotspots' for threatened



Fig 1. Study region. South Australia on the mainland of the Australian continent (plus its off-shore islands), showing the proportion of remnant native vegetation within a moving window of 25 x 25 km (large areas with no vegetation in the north are salt lakes). Scale bar: 200 km. Inset: context within Australia.

doi:10.1371/journal.pone.0144779.g001

species (see http://www.environment.gov.au/biodiversity/conservation/hotspots/nationalbiodiversity-hotspots, accessed 14 September 2015) and the regional trajectory of species (see http://www.environment.sa.gov.au/managing-natural-resources/plants-and-animals/ Threatened species ecological communities/Regional significant projects/Regional Species Conservation Assessment Project, accessed 14 September 2015).

Biodiversity sampling issues

Biodiversity mapping is an empirical exercise in representing species inventory data. However, given that large-scale biodiversity mapping studies typically use data from existing sources such as herbarium or museum records collected non-systematically and for a different purpose, biases in the underlying data may influence the results [13,21]. Primary sampling issues for biodiversity mapping include: 1. uneven and under- sampling, resulting in biased richness estimates; and 2. that many metrics are correlated with *species richness*. These factors can make it difficult to disentangle metrics from sampling intensity and *species richness*. The absence of information for un-sampled areas is also a major issue for poorly sampled taxa and regions [13]. However, this is less of an issue for this study because, between herbarium and Biological Survey data, the vascular flora of South Australia is relatively well sampled, a significant legacy put to use here.

Plot-based data recording species according to systematic effort criteria make it possible to use the accumulation of species to estimate actual *species richness* from that observed, for example via rarefaction or non-parametric estimators [3,22-24]. However, for the purposes of identifying relative patterns of biodiversity, it is not always necessary to correct metrics for

sampling: it may be more relevant to ask whether metrics are higher than expected, given the context of sampling.

Aims

We aimed to identify centres of high vascular plant biodiversity in South Australia by compiling existing datasets and mapping a suite of quantitative biodiversity metrics (<u>Table 1</u>), making use of significant yet under-utilised data resources. Having identified centres of biodiversity, we aimed to overlay spatial information on associated threats at a similar scale for context. The conservation issues considered were sensitivity to climate change as predicted from spatial analogues, exposure to habitat fragmentation, introduced species and fire regimes.

Materials and Methods

Study area

Geophysical context. South Australia makes up a sub-continental land area of 983,482 km², comprising approximately 13% of the Australian continent, and consisting predominantly of extensive arid (mean annual rainfall approximately 100–300 mm) plains in the north and low-relief mediterranean-climate (mean annual rainfall approximately 300–1000 mm) regions in the south (source: Atlas of South Australia (http://www.atlas.sa.gov.au), accessed 12 November 2014; Bureau of Meteorology (http://www.bom.gov.au), accessed 12 November 2014). Major landscape features include large salt lakes (the largest, Lakes Gairdner, Eyre, Frome and Torrens), and isolated mountain ranges to 1400 m ASL in the north-west (Central Ranges IBRA bioregion) and to 1100 m ASL along the Mount Lofty–Flinders Ranges (Flinders-Lofty Block and Kanmantoo IBRA bioregions). The state's coast is characterised by three peninsulas (from west to east: Eyre, Yorke, Fleurieu) and off-shore islands, including Kangaroo Island, Australia's third largest, at 4,405 km² (Fig A in <u>S1 Appendix</u>).

Vegetation condition. Eighty-seven percent of the state's land area is arid and retains 96% of its native vegetation, although there is extensive use as rangelands [6,25]. The remaining 13% of land area in the mediterranean-climate south retains 4–26% of native vegetation and, although broad-acre clearing has been prohibited since 1991, illegal and legal clearing of degraded vegetation continues [6].

Despite progressive increases to the area of land under formal conservation protection, vegetation condition in many areas is in decline due to altered fire regimes, introduced pests and poor recruitment [6,26]. Management of land-use in remnants is also an on-going issue. Of native vegetation sites in the Northern and Yorke Natural Resource Management region, for example, 60% have poor control of livestock grazing [6]. In the Adelaide–Mount Lofty Ranges region there has been peri-urban land-use intensification, such as the conversion of lightly grazed paddocks to vineyards [25].

Datasets

Species data. A set of 330,004 individual records of 3,083 vascular plant species from 14,328 systematically surveyed field plots across the state was obtained from the *Biological Survey of South Australia*, or 'BSSA' [27,28]. BSSA plots were established to systematically sample and document variation in ecological communities in the context of their environments to provide information for reserve planning and the identification of areas with high biological significance [10], although such an analysis has never been conducted at state level. The standard plot size was 30 x 30 m, although some plots were larger in arid areas, and some earlier plots were 10 x 10 m.

	ONE
--	-----

Metric (full)	Metric (short)	Herbarium	Plots	Combined data	Correction or test
Species richness of:					
All species		х	х		Non-parametric estimator
Native species		х	х		-
 Introduced species 			х		-
Categorical endemics		х	х		Compare observed ~ expected; non-parametric outlier statistics
 Conservation-dependent species 		х	х		-
Georeferenced weighted endemism	GWE		x		Compare observed ~ expected; non-parametric outlier statistics
Georeferenced phylogenetic endemism	GPE		х		Compare observed ~ expected; non-parametric outlier statistics
Phylogenetic diversity	PD		x		Compare observed ~ expected; non-parametric outlier statistics
Sørensen dissimilarity, mean of all pairwise comparisons				x	•
Sørensen dissimilarity, mean of comparisons among 5x5 cell moving window				x	-
Correspondence Analysis	CA			x	-
Canonical Correspondence Analysis	CCA			x	Mean difference along constrained axis within 3x3 moving window
Canonical Correspondence Analysis	CCA			x	Slope of local polynomial regression to estimate high turnover and sensitivity
Hierarchical classification analysis	HCA			x	Replacement component of Sørensen dissimilarity only; selected metrics re-calculated for 40 vegetation groups

Table 1. Biodiversity metrics mapped for South Australia and associated datasets and tests.

doi:10.1371/journal.pone.0144779.t001

BSSA data were supplemented with plot-based data from the Terrestrial Ecosystem Research Network's (TERN) 'AusPlots Rangelands' [29]. Data were subsetted to plant species vouchered following exhaustive visual searches of 27 one hectare plots in the arid zone of South Australia, which is generally under-sampled, resulting in an additional 967 records of 231 species [30].

In addition to these datasets, herbarium records were obtained through Australia's Virtual Herbarium with the following filters: state of South Australia as the spatial extent; records from the State Herbarium of South Australia; spatially valid records; no uncertain specimen determinations; identified to species or lower taxonomic level [31]. Records for genera previously belonging to family Chenopodiaceae (now Amaranthaceae) were manually extracted, as the above filters excluded them due to missing classification fields. The resulting dataset consisted of 396,446 records of 4,501 species.

Fields associated with species names in the South Australia census of vascular plants [32; electronic version updated June 2014] were matched with species records to subset data to native or introduced species and species considered endemic to South Australia (updated 2014, P.J.Lang pers. comm.).

Environmental and fire history data. The following map layers were obtained via 'Map-Land' (Department of Environment, Water and Natural Resources, SA): polygons of extant native vegetation across South Australia (for calculation of habitat fragmentation indices); fire history layers consisting of polygons of fire events. Raster maps of bioclimatic variables were obtained using tiles from WorldClim [33] trimmed to the extent of South Australia.

Mapping resolution

Biodiversity was mapped over 0.25° grid cells, which was found to be a reasonable balance between high enough resolution for interpretation and a low enough resolution to capture a representative sample of species records. Biodiversity rasters were subsequently smoothed at a resolution of approximately 1 km² grid cells (representing values for the surrounding area of 0.25°). Smoothing at a finer resolution allowed areas outside of the coastline to be trimmed and provides a more realistic output that minimises artificial differences between adjacent cells, although mainly visual. We recognise that at this scale there is likely to be variation within grid cells in levels of biodiversity and vegetation remnancy, for example.

Sampling corrections

Sampling issues included uneven intensity, biases in locations and taxa and correlation between *species richness* and other metrics. In some instances, we calculated raw metrics, with only qualitative interpretation of sampling influence. This was the case for herbarium data which, despite in creased taxonomic and spatial coverage over plots, are more biased towards highways and populations centres, have fewer records representing occurrences of common species and are more problematic to correct for sampling intensity [34–36]. For total *species richness* estimated from plot data, we employed the bootstrap non-parametric richness estimator to boost estimates of richness in under-sampled grid cells [37,38]. Non-parametric estimators were chosen among alternatives such as rarefaction as they are computationally efficient, although we recognise these estimates are likely still biased in cells with very low sampling [23].

For metrics associated with endemism or phylogenetic diversity, we used non-parametric tests, described below along with the specific metrics, as to whether observed values were different to that expected, given the number of species recorded, a method that accounts for the influence of both under-sampling and *species richness* itself [27].

For selected metrics, we calculated a 3 x 3 cell moving window mean to smooth artificial distinctions between adjacent cells due to spatial errors or sampling [13]. The moving window provides estimates for un-sampled grid cells based on the assumption that they will be similar to their immediate neighbours, although we acknowledge this may occasionally produce anomalies.

Biodiversity metrics

Species metrics. Species richness was calculated using plot-based data for all species; native species only; introduced weed species only, conservation-dependent species only (defined as those species with a rating under the South Australian National Parks and Wildlife Act 1972) and known South Australian endemics only (*endemic richness*). Herbarium records were used to map occurrences of Buffel Grass (*Cenchrus ciliaris* L.), a weed previously identified as having a disproportionate impact in ecosystem function and native species diversity in arid Australian ecosystems [39,40].

Endemic richness was compared to the null model that presences of endemic species are random, given observed *species richness*, and do not represent any special pattern. The aim was to distinguish concentrations of endemic species from areas of higher *species richness* that, by default, may have higher absolute numbers of endemics. Observed *endemic richness* species was compared to that in 1,000 replicate random draws of species from the overall pool (with sampling probabilities weighted by observed species frequencies), and calculating nonparametric two-tailed statistical significance (considered statistically significant if lying within or beyond the upper or lower 2.5%), and a continuous outlier metric (the factor of the interquartile range by which observed values out-lie the median of the null distribution). *Georeferenced weighted endemism* (GWE: *species richness* weighted by restriction in extent of occurrence; [41]) was mapped using the implementation of Guerin *et al.*, [27] with the distance spanning species occurrences as weights. Raw scores were compared to 1,000 replicated random draws (null distribution) to calculate statistical significance and categorical and continuous outlier metrics as for *endemic richness* (method also described in Guerin *et al.*, [27]). GWE included native species only, as geographically restricted introduced species are not regarded as having conservation value.

Phylogenetic metrics. A phylogenetic tree of all plant species in the dataset was generated from Phylomatic Version 3 (<u>http://phylodiversity.net/phylomatic/</u>) and tree R20120829 [42]. Node age constraints were applied after Wikström *et al.*,[43] along with branch-length adjustment using the bladj algorithm, a simple estimator that spaces nodes evenly between age constraints [44].

Phylogenetic diversity (PD; [45]) was calculated by summing the branch-lengths of the phylomatic tree after pruning to species occurring in each grid cell. *Georeferenced phylogenetic endemism* (GPE: phylogenetic diversity weighted by restriction in the extent of occurrence of branches; [41,46] was calculated using the implementation of Guerin & Lowe [47] with the maximum distance spanning the geographical range of branches for weights.

Raw PD and GPE scores were tested as to whether they were higher or lower than expected, given observed *species richness*, with a non-parametric method equivalent to that described for *endemic richness* and outlined for phylogenetic metrics in Guerin & Lowe [47].

Turnover analysis

Datasets. Species records from plot and herbarium data were combined to obtain the most complete possible representation of species composition (as opposed to unbiased estimates of *species richness* or other alpha biodiversity metrics). With these records, we generated a matrix of species presence/absence in grid cells for beta diversity analysis.

Beta diversity and climate sensitivity. We calculated pairwise Sørensen dissimilarities for grid cells. As a measure of compositional uniqueness in the context of the whole study region, mean dissimilarities for each grid cell compared to all other cells were mapped, excluding cells with fewer than 20 observations, for which dissimilarities were inflated. As a measure of more local heterogeneity, mean dissimilarities were also calculated among all cells within a 5 x 5 cell moving window.

We applied Correspondence Analysis (CA) to the occurrence matrix and mapped out scores for the first axis. The first CA axis is frequently reported, and visually appeared in this case, to correlate with major latitudinal climatic gradients. We explored this further with scatterplots and calculation of correlation coefficients (using Spearman's rho because the relationships appeared monotonic but obviously non-linear) for axis scores and bioclimatic gradients. The variable 'mean temperature of the hottest month' was this most highly correlated with axis scores ($\rho = -0.92$), followed by 'mean annual rainfall' ($\rho = 0.81$). We therefore repeated the ordination analysis using Canonical Correspondence Analysis (CCA) constrained by 'mean temperature of the hottest month'. Subsequently, we calculated the mean distance along the resulting CCA axis among a moving window of 3 x 3 cells to highlight areas where composition is changing more rapidly along this axis, and used local polynomial regression ('loess') to explore non-linear relationships between weighted average scores (based on species scores for sites and not constrained to be linear combinations of constraining variables) and this gradient. Regions with the highest slope (change in axis score over change in temperature variable) were interpreted as having a species composition more sensitive to spatial changes in temperature and expected temporal changes [7,48]. CCA has been shown to be a robust method because, unlike CA, it is not prone to artefacts such as the arch effect [48, 49].

Hierarchical classification

Species composition was used to classify major vegetation groups based on hierarchical clustering with complete linkage agglomeration and Sørensen replacement (not richness-difference) dissimilarities [50]. Grid cells were excluded from training the classification if there were fewer than 20 records within them, to avoid biases and difficulty classifying cells.

The classification was cut into 40 groups, which, although arbitrary, gave a reasonable number of evenly sized groups. Groups were mapped onto a raster layer with a simple smoothing function to account for under-sampled grid cells by assigning the most frequent vegetation group within a 3 x 3 moving window. The five most frequent species in constituent grid cells were identified for each group.

We matched the classification to GPE to represent the biodiversity value of the 40 vegetation groups because GPE is increased by elements of *species richness*, range restriction and phylogenetic diversity. Cells in each group were assigned the mean GPE score. Secondly, to represent vegetation groups or habitats impacted by weeds, we calculated the richness of introduced species recorded within each group.

Habitat fragmentation and fire layers

We converted remnant native vegetation polygons to a $\sim 1 \text{ km}^2$ resolution raster, and for each grid cell calculated the proportion of native vegetation within a 25 x 25 km moving window, to represent the fragmentation level of the habitat matrix surrounding each location. The aim was to identify the overall remnancy level at coarse scale to identify heavily cleared and fragmented landscapes, whereas finer resolution mapping would be needed to accurately determine habitat configuration at more local scales. A map of fire frequency was generated and rasterised then used to visually identify the intersection of fire prone ecosystems with high biodiversity. There was no intention to analyse whether fire regimes were appropriate from a biodiversity perspective, the aim was simply to identify whether fire management interacted with any identified biodiversity centre.

Software

Analyses were conducted with custom scripts in base R [51], and using functions from packages including 'ape', 'vegan', 'raster', 'labdsv', 'maptools', 'simba', 'plyr' and the functions of Guerin *et al.*, [27] and Guerin & Lowe [47]. The species list for phylomatic was prepared with a custom script that attached family names via automated searches of the Atlas of Living Australia (www.ala.org.au) and The Plant List (www.theplantlist.org).

Results

Sampling intensity

A map of the spatial density of herbarium collections revealed high spatial coverage but a significant bias to locations near the capital city of Adelaide (Fig B in <u>S1 Appendix</u>). The spatial coverage of biological survey plots was less complete but more even in intensity.

Biodiversity metrics

Species richness was generally higher throughout the southern agricultural areas than in the arid north (Fig.2a). The Mount Lofty Ranges, east of Adelaide, had very high *species richness*



Fig 2. Maps of measured biodiversity metrics. Maps are smoothed at a resolution of 1 km² with values given per surrounding 0.25° x 0.25° area: (a) Estimated *species richness* based on non-parametric estimator (plot data); (b) *georeferenced weighted endemism* of native species (plot data)–continuous outlier metric; (c) *phylogenetic diversity* (plot data)–continuous outlier metric; (d) *georeferenced phylogenetic endemism* (plot data)–continuous outlier; (e) *species richness* of conservation-dependent species (plots); (f) *species richness* of categorical South Australian endemics (plot data).

PLOS ONE

with herbarium data coinciding with large numbers of collections in this region (Figs B and C in <u>S1 Appendix</u>).

Conservation-dependent *species richness* differed according to dataset. Herbarium data suggested Adelaide-Mount Lofty Ranges and Lower South East were the highest, followed by western Kangaroo Island, southern Flinders Ranges, and lower Eyre Peninsula. Plot data suggested a more even richness among those locations, but lower richness in the Lower South East (Fig 2e and Fig C in <u>S1 Appendix</u>).

Concentrations of (categorical) *endemic richness* were evident on Adelaide-Mount Lofty Ranges, Kanagaroo Island, lower Eyre Peninsula and scattered areas of the Flinders Ranges. The highest absolute numbers, and the most outlying and statistically significant concentrations, were on lower Eyre Peninsula and Kangaroo Island (Fig 2f and Fig D in S1 Appendix.

GWE was higher than expected in several regions: Adelaide–Mount Lofty Ranges, western Kangaroo Island, Lower South East, and two locations in the far north-west of the state: 1. The northern (ranges) section of the APY (Anangu Pitjantjatjara Yankunytjatjara) lands, near the northern state border; 2. The Mamungari Conservation Park, near the western state border (Fig 2b and Fig E in <u>S1 Appendix</u>). GWE was statistically, but barely, higher than expected for the lower Eyre Peninsula. A large area across the middle of the state had slightly lower than expected endemism.

Raw PD had a similar pattern to corrected *species richness* with higher values in southern agricultural regions (Fig F in <u>S1 Appendix</u>). When compared to null expectations, phylogenetic diversity was statistically—but barely—higher than expected in many locations in the south (<u>Fig 2c</u> and Fig F in <u>S1 Appendix</u>). PD was above median expectations in the north-west but this was not statistically significant. A large area of the arid interior had slightly lower than expected phylogenetic diversity (statistically significant).

GPE was concentrated, and higher than expected, in the same locations as for GWE, although with higher relative scores for the north-west (Fig 2d and Fig G in <u>S1 Appendix</u>). These areas were statistically significant, along with a small number of other areas that were only slightly above expected values. Large areas mostly through the middle of the state had slightly lower than expected phylogenetic endemism.

Turnover analysis

Beta diversity based on state-wide dissimilarity appeared to be biased for under-sampled grid cells, as the poorly sampled areas scored higher than those with better access (Fig H in <u>S1</u> <u>Appendix</u>). Notwithstanding, the general pattern was scores that were low for the arid zone and increased to the south. Moving-window analysis revealed the opposite pattern, whereby the southern agricultural areas had relatively low beta diversity, whereas there was high beta diversity across the entire arid zone in the north (Fig H in <u>S1</u> <u>Appendix</u>).

Correspondence analysis returned a primary compositional axis running approximately north–south through the state (Fig H in <u>S1 Appendix</u>). When this ordination was repeated constrained by the 'mean maximum temperature of the hottest month' climate variable in a CCA analysis, change along the constrained axis within a moving spatial window showed an area overlapping the boundary between arid and mediterranean regions of the state where turnover was higher (Fig 3). The largest rate of change in the compositional axis modelled along the temperature gradient also corresponded to this ecotone (Fig 4).

Classification analysis

Classification analysis identified 40 specified vegetation clusters and their most frequent species (Fig I and Table A in <u>S1 Appendix</u>). Mean GPE for grid cells assigned to the clusters showed



Fig 3. Beta diversity. Here beta diversity is represented by the mean change in a Canonical Correspondence Analysis axis coordinate within a moving window of 3 x 3 grid cells, when this axis is constrained by 'mean maximum temperature of the hottest month' (combined data). Higher values represent more rapid turnover along the compositional axis.

that high scoring clusters were restricted to southern coastal areas of the state. Clusters with the highest GPE were found on Kangaroo Island–Fleurieu Peninsula(–south-east) followed by the lower Yorke Peninsula–Adelaide-Mt Lofty Ranges cluster.

Exposure to habitat fragmentation, fire and invasive species

Fragmentation mapping clearly showed a dichotomy in land-uses, with the arid north nearly completely vegetated, whereas the southern areas have been reduced to 0-30% vegetation (Fig 1). Highly fragmented landscapes coincided with most, but not all, areas of high biodiversity.

Fire mapping highlighted areas prone to higher fire frequency (Fig 5). While most patches of remnant native vegetation in higher rainfall areas of the state have been subject to recorded fires, high fire frequency areas of note include the APY lands (NW South Australia) and western Kangaroo Island. Low fire frequency areas generally corresponded to vegetation that is either too sparse or fragmented to carry significant fires.

Richness of introduced species based on plots (which sampled remnant vegetation) was concentrated in agricultural areas close to population centres, most notably high from the Fleurieu Peninsula north to the southern Flinders Ranges and to a lesser extent southern Eyre and York Peninsulas, while Kangaroo Island and the south east of the state were notably lower (Fig 6 and Fig C in S1 Appendix). Weed richness was high (i.e. \sim >200 spp.) for several



Fig 4. Identified centres of plant biodiversity. The base raster map shows the proportion of remnant vegetation in a moving window (see Fig 1). Contours show the rate of change in species composition with spatial increases in temperature (predicted from a locally weighted nonlinear regression of the first CCA axis), where more negative slopes indicate higher climate sensitivity. Circles show locations of 6 centres referred to in Table 2. See Table 2 for more detail on the identified centres.

vegetation clusters covering diverse areas such as the Nullarbor Plain (western coast-line), western Eyre Peninsula and lower Yorke Peninsula, while clusters in the arid zone had few weed species (i.e. $\sim <<100$ spp.), excepting the northern Flinders Ranges and the dunefields of the Great Victoria Desert bioregion, which had medium weed richness (i.e. $\sim 100-150$ spp.; Fig I in <u>S1 Appendix</u>). Recorded Buffel Grass occurrences were concentrated in the far north-east and north-west of the state as well as in the vicinity of major transport corridors to the capital city of Adelaide in the south (Fig.6).

Centres of biodiversity

We identified six locations as centres of biodiversity (in no particular order; <u>Table 2</u>; <u>Fig 4</u>): **1**. Western Kangaroo Island; **2**. Southern Mount Lofty Ranges; **3**. APY Lands (NW South Australia); **4**. Southern Flinders Ranges; **5**. Southern Eyre Peninsula; **6**. Lower South East. Conservation issues associated with these locations were determined via the maps of climate sensitivity, habitat fragmentation, weed diversity and fire frequency (<u>Table 2</u>).



Fig 5. Number of recorded significant fires. Base map shows IBRA bioregions (Fig A in <u>S1 Appendix</u>). Numbers show locations of 6 centres referred to in <u>Table 2</u>.

Discussion

We set out to provide basic biodiversity information collectively on some 4,500 species of vascular plants across South Australia. This analysis of species occurrences used intensively sampled biological data and a suite of sophisticated, objective metrics to map areas of high diversity. Six localised areas of the state were clearly highlighted (Fig 4). Although neither conservation prioritisation per se nor detailed assessment of ecosystem condition or trajectory were our primary aim, mapping of major conservation issues showed that each biodiversity centre is associated with issues such as habitat fragmentation and weed invasion at coarse scales.

The southern Mount Lofty Ranges scored highly for unique species and overall high diversity, but has been subjected to the most disturbance since European settlement, with high levels of weed invasion and habitat fragmentation, which suggests a higher resolution assessment is warranted. The western side of Kangaroo Island has perhaps the most significant plant biodiversity in South Australia, scoring consistently highly, but has higher reservation levels and lower incidences of weed species.

This study was made possible by the wealth of information on the distribution of species within the state including intensive herbarium sampling and a Biological Survey program that has systematically recorded vegetation across the state. These data allowed the use of numerical methods highlighting different aspects of biodiversity that can be independently repeated, without reliance upon expert opinion or modelling to fill gaps.



Fig 6. Species richness of introduced plant species (plot data). Circles represent the density of individual records of Buffel Grass (*Cenchrus ciliaris* L.) within 1° grid cells on a logarithmic scale. Note, the number of records is influenced by sampling biases and does not directly indicate abundance. Numbers show locations of centres referred to in <u>Table 2</u>.

doi:10.1371/journal.pone.0144779.g006

Comparison of metrics

Crisp *et al.*, [17] concluded that the Adelaide–Kangaroo Island region of South Australia had high endemism at a continental scale. In our study, areas highlighted for endemism depended

Table 2. Identified centres of plant biodiversity in South Australia.

Region	Biodiversity assets	Associated conservation issues
1.Western Kangaroo Island	1. Higher than expected GWE and GPE; 2. Concentrations of conservation- dependent species and categorical endemics; 3. High <i>species richness</i>	1. High fire frequency
2. Southern Mount Lofty Ranges	1. Higher than expected GWE and GPE; 2. Concentrations of conservation- dependent species; 3. High <i>species richness</i>	1. High levels of habitat fragmentation; 2. Many introduced species
3. APY lands (NW South Australia)	1. Higher than expected GWE and GPE	1. High fire frequency; altered fire regimes; 2. Management of Buffel Grass invasion
4. Southern Flinders Ranges	1. Concentrations of conservation-dependent species; 2. High species richness	1. High grazing disturbance levels; 2. High climate sensitivity
5. Southern Eyre Peninsula	1. Concentrations of conservation-dependent species and categorical endemics; 2. High species richness	1. High levels of habitat fragmentation; 2. Many introduced species
6. Lower South East	1. Higher than expected GWE and GPE; 2. Concentrations of conservation- dependent species (herbarium data)	1. High levels of habitat fragmentation

doi:10.1371/journal.pone.0144779.t002

somewhat on the method. GWE highlighted the Lower South East and north-west of South Australia in addition to Kangaroo Island and Adelaide-Mount Lofty Ranges. The former locations are geographically unique within South Australia and lie in the corners of political borders. By comparison, categorical *endemic richness* highlighted Kangaroo Island and lower Eyre Peninsula in particular. Both metrics convey relevant information for conservation in a state context: GWE for highlighting species with restricted ranges in the state (although potentially biased by distributions overlapping state borders); *endemic richness* for highlighting species only found in the state (although a function of political boundaries and potentially biased by species with restricted ranges occurring in the middle of the state being more likely endemic to it).

The APY lands have areas of topographical variation in the Tomkinson, Mann, Musgrave, Everard, and Indulkana Ranges and around granite outcrops, providing refugia such as gorges and high altitude summits [52]. The GWE and GPE results highlight the value of conservation management in the northern APY lands. It is already recognised that fire regimes and the spread of invasive species including feral animals and Buffel Grass need to be managed in partnership with traditional owners to promote ecological outcomes [26,53]. We are cautious about the high GPE scores immediately south of the APY lands within the Mamungari Conservation park (Great Victoria Desert bioregion) because sampling has been limited by access, and in any case, threat levels and therefore need for management are relatively low given the park's UNESCO World Biosphere Reserve status.

Beta diversity highlighted no particular location, and which general region scored higher depended on whether comparisons were state-wide or local. We interpret the high heterogeneity of species composition along a major ordination axis across the region separating the arid and mediterranean biomes as an ecotone [48].

Associated conservation issues

Climate sensitivity. High spatial turnover in species composition along a major latitudinal climatic gradient, as evident in our data across an ecotone region, can be interpreted as potential high sensitivity to temporal climate change [54]. Space-for-time substitution of course assumes that the spatial sensitivity of species composition to temperature will translate to sensitivity to temporal changes in climate. Mean daily summer maximum temperatures across this region are predicted to rise by two degrees this century over 1986–2005 levels, based on median predictions of 15 models under an intermediate emissions pathway [7]. This is significant because a spatial difference of two degrees is associated with significant shifts along an associated species composition axis.

All but one of the identified centres of biodiversity, the southern Flinders Ranges, occur outside of the area with the highest climate sensitivity, but climate adaptation may be a major concern for ecosystem management in this zone and a consideration for less sensitive areas. It is important to note that there are other axes of compositional variation that may not respond at all to climate change, and that the translation of spatial changes in vegetation along climatic gradients to temporal change is an assumption that needs to be tested through long term monitoring.

Habitat fragmentation. The dichotomy between extensive and intensive land-uses between the state's arid/mediterranean biomes presents contrasting management issues among the identified biodiversity centres. The lower Eyre Peninsula, Adelaide-Mount Lofty Ranges and Lower South East centres have been the most extensively cleared, suggesting addressing the impacts of habitat fragmentation may be an important concern in these areas. In contrast, extensive land management issues such as invasive species and fire regimes are critical for the APY lands, where vegetation condition rather than remnancy is the key issue. The mapping of habitat presence was relatively coarse at a resolution of 1 km^2 . A future extension of this study would be to map biodiversity metrics and landscape characteristics, including habitat remnancy and configuration at much higher resolution over identified biodiversity centres.

Fire regimes. The most extensively fire prone region according to frequency mapping is the APY lands. Given low levels of habitat fragmentation, maintaining appropriate fire regimes across this area is a primary concern for conservation. Fire management is also relevant to managing remnant vegetation in the south of the state, where there are localised areas of high fire frequency, notably western Kangaroo Island. Our mapping recorded fire frequency but not intensity or season, nor how appropriate fire regimes have been for biodiversity conservation to date. Future extensions to work would need to assess both the most appropriate fire regimes for maintaining biodiversity but also how fire regimes may be changing. The present study gives a broad-brush indication of the main areas in which fire is an issue.

Invasive species. Introduced *species richness* was higher in southern agricultural areas receiving higher rainfall and in major population corridors and this may translate to pressure on the maintenance of native species. A notable exception is Kangaroo Island, which has relatively low weed richness despite a mesic climate, perhaps due to isolation from mainland propagule sources and lower levels of habitat fragmentation.

Weed richness does not account for abundance or directly provide information on whether weed diversity correlates negatively with native diversity. Particular introduced weeds may disproportionately contribute to conservation impacts. Buffel Grass, in particular, has been shown to invade relatively unperturbed habitats and to have measurable impacts on native species diversity and fire regimes [39,40,55,56]. The major impact zone for Buffel Grass at present is the north-west of the state, where feedbacks with fire regimes are an issue.

Conclusions

We identified six centres of high plant biodiversity across South Australia, based on metrics mapped from plot and herbarium data. We also mapped their spatial interaction with climate sensitivity, habitat fragmentation, fire frequency and weed *species richness*. Areas highlighted were not always congruent among metrics or datasets, which affirms our approach of using a suite of complementary metrics and alternative data sources to increase confidence. Herbarium data captured more species and locations but were more uneven in intensity. Highlighted centres of endemism differed between categorical and weighted metrics, perhaps due to contrasting biases caused by political boundaries.

Species composition in the arid-mediterranean ecotone was most sensitive to climate, although only one biodiversity centre—the southern Flinders Ranges—was located in this zone. Three of the biodiversity centres coincided with highly fragmented vegetation, and four with high weed diversity. For the APY lands, management of fire and Buffel Grass were the main associated issues.

Practical application of these results would benefit from higher resolution assessments within identified biodiversity centres and information on the cost:benefit ratio of intervention, taking into account potential biodiversity/area returns but also costs such as land purchase, lost production opportunity and the market price for management actions.

Supporting Information

S1 Appendix. Additional maps and tables of biodiversity metrics. (PDF)

Acknowledgments

We thank Ben Sparrow, Andrew Tokmakoff, Anita Smyth, Phil Pisanu, Damon Ezis, Michelle Waycott, Susan Sweeney, Peter Lang, John McDonald, Sandy Carruthers, Dan Rogers and MapLand. Public data were obtained via the ÆKOS Data Portal (<u>http://www.portal.aekos.org.</u> au/) at TERN Eco-informatics, and Australia's Virtual Herbarium (AVH; <u>http://avh.chah.org.</u> au).

Author Contributions

Conceived and designed the experiments: GRG EB ZB AJL. Performed the experiments: GRG EB ZB. Analyzed the data: GRG EB ZB. Wrote the paper: GRG EB ZB AJL. Designed the software used in analysis: GRG.

References

- 1. Wilson KA, Underwood EC, Morrison SA, Klausmeyer KR, Murdoch WW, Reyers B, et al. Conserving biodiversity efficiently: what to do, where, and when. PLoS Biol. 2007; 5: e223. PMID: <u>17713985</u>
- 2. Eken G, Bennun L, Brooks TM, Darwall W, Fishpool LD, Foster M, et al. Key biodiversity areas as site conservation targets. Bioscience. 2004; 54: 1110–8.
- Reese GC, Wilson KR, Flather CH. Performance of species richness estimators across assemblage types and survey parameters. Glob Ecol Biogeogr. 2014; 23: 585–94.
- Guerin GR, Martín-Forés I, Biffin E, Baruch Z, Breed MF, Christmas MJ, et al. Global change community ecology beyond species-sorting: a quantitative framework based on mediterranean-biome examples. Glob Ecol Biogeogr. 2014; 23: 1062–72.
- 5. Gill AM, McKenna DJ, Wouters MA. Landscape fire, biodiversity decline and a rapidly changing milieu: a microcosm of global issues in an Australian biodiversity hotspot. Land. 2014; 3: 1091–136.
- 6. EPA. State of the Environment South Australia. Adelaide, South Australia; 2013.
- Charles SP, Fu G. Statistically downscaled climate change projections for South Australia. Adelaide, South Australia; 2015.
- McCallum KP, Guerin GR, Breed MF, Lowe AJ. Combining population genetics, species distribution modelling and field assessments to understand a species vulnerability to climate change. Austral Ecol. 2014; 39: 17–28.
- 9. Thackway R, Cresswell ID. An interim biogeographic regionalisation for Australia: a framework for establishing the national system of reserves. Canberra; 1995.
- Robinson AC, Casperson KD, Canty PD, MacDonald CA. A biological survey of the Gawler Ranges, South Australia in October 1985. Adelaide: National Parks and Wildlife Service, Department of Environment and Planning, South Australia & South Australian Museum, Department for the Arts, South Australia, D.J. Woolman, Government Printer; 1988.
- 11. Slatyer C, Rosauer D, Lemckert F. An assessment of endemism and species richness patterns in the Australian Anura. J Biogeogr. 2007; 34: 583–96.
- González-Orozco CE, Laffan SW, Miller JT. Spatial distribution of species richness and endemism of the genus Acacia in Australia. Aust J Bot. 2011; 59: 601–609.
- Stevenson LA, González-Orozco CE, Knerr N, Cargill DC, Miller JT. Species richness and endemism of Australian bryophytes. J Bryol. 2012; 34: 101–7.
- Mishler BD, Knerr N, González-Orozco CE, Thornhill AH, Laffan SW, Miller JT. Phylogenetic measures of biodiversity and neo- and paleo-endemism in Australian Acacia. Nat Commun. 2014; 5: 4473. doi: <u>10.1038/ncomms5473</u> PMID: <u>25034856</u>
- González-Orozco CE, Brown AHD, Knerr N, Miller JT, Doyle JJ. Hotspots of diversity of wild Australian soybean relatives and their conservation in situ. Conserv Genet. 2012; 13: 1269–81.
- González-Orozco CE, Thornhill AH, Knerr N, Laffan S, Miller JT. Biogeographical regions and phytogeography of the eucalypts. Divers Distrib. 2014; 20: 46–58.
- 17. Crisp, Laffan, Linder, Monro. Endemism in the Australian flora. J Biogeogr. 2001; 28: 183–98.
- Laffan SW, Crisp MD. Assessing endemism at multiple spatial scales, with an example from the Australian vascular flora. J Biogeogr. 2003; 30: 511–20.

- Byrne M, Yeates DK, Joseph L, Kearney M, Bowler J, Williams MAJ, et al. Birth of a biome: insights into the assembly and maintenance of the Australian arid zone biota. Mol Ecol. 2008; 17: 4398–417. doi: <u>10.1111/j.1365-294X.2008.03899.x</u> PMID: <u>18761619</u>
- Traill LW, Wanger TC, de Little SC, Brook B. Rainfall and temperature variation does not explain arid species diversity in outback Australia. Res Reports Biodivers Stud. 2013; 3: 1–8.
- Droissart V, Hardy OJ, Sonké B, Dahdouh-Guebas F, Stévart T. Subsampling herbarium collections to assess geographic diversity gradients: a case study with endemic Orchidaceae and Rubiaceae in Cameroon. Biotropica. 2012; 44: 44–52.
- Colwell RK, Coddington JA. Estimating terrestrial biodiversity through extrapolation. Philos Trans R Soc London B. 1994; 345: 101–18.
- Clarke K, Lewis M, Ostendorf B. Additive partitioning of rarefaction curves: Removing the influence of sampling on species-diversity in vegetation surveys. Ecol Indic. 2011; 11: 132–9.
- Cardoso P, Rigal F, Borges PA V., Carvalho JC. A new frontier in biodiversity inventory: a proposal for estimators of phylogenetic and functional diversity. Faith D, editor. Methods Ecol Evol. 2014; 5:452– 61.
- EPA. The State of our Environment: State of the Environment report for South Australia 2008. Adelaide, South Australia; 2008.
- Paltridge R, Latz P, Pickburn A, Ward M, Ward J, Davey M. Working with Anangu to conserve rare plants in the Anangu Pitjantjatjara Yankunytjatjara Lands of South Australia. Australas Plant Conserv. 2011; 19: 10–1.
- Guerin GR, Ruokolainen L, Lowe AJ. A georeferenced implementation of weighted endemism. Methods Ecol Evol. 2015; 6: 845–852.
- 28. Department of Environment Water and Natural Resources. Biological Survey of South Australia—Vegetation Survey, Biological Database of South Australia (endemism mapping subset). Version 1.0. Adelaide, South Australia; 2014. 10.4227/05/5444BAF32E94C. Obtained from Australian Ecological Knowledge and Observation System Data Portal (ÆKOS, <u>http://www.portal.aekos.org.au/</u>), made available by State of South Australia (Department of Environment, Water and Natural Resources), Adelaide, South Australia. Accessed 20 October 2014 (<u>http://portal.aekos.org.au/dataset/168004</u>).
- Tokmakoff A, Sparrow B, Turner D, Lowe A. AusPlots Rangelands field data collection and publication: infrastructure for ecological monitoring. IEEE 10th International Conference on e-Science. 2014; 249– 55.
- 30. TERN AusPlots. Terrestrial Ecosystem Research Network AusPlots—Ausplots Rangelands Survey Program (biodiversity mapping supplement/subset), Version 1.0. 2015. 10.4227/05/54C1B45A4CF2F. Obtained from Australian Ecological Knowledge and Observation System Data Portal (ÆKOS, <u>http://www.portal.aekos.org.au/</u>), made available by University of Adelaide. Accessed 23 January 2015. (<u>http://portal.aekos.org.au/</u>dataset/172373)
- **31.** AVH. Australia's Virtual Herbarium. Council of Heads of Australiasian Herbaria. 2014. Available from: <u>http://avh.chah.org.au</u>, accessed 31 October 2014.
- Barker WR, Barker RM, Jessop JP, Vonow HP, editors. Census of South Australian vascular plants. 5th ed. Adelaide: Botanic Gardens of Adelaide & State Herbarium; 2005.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. Very high resolution interpolated climate surfaces for global land areas. Int J Climatol. 2005; 25: 1965–78.
- Geri F, Lastrucci L, Viciani D, Foggi B, Ferretti G, Maccherini S, et al. Mapping patterns of ferns species richness through the use of herbarium data. Biodivers Conserv. 2013; 22: 1679–90.
- Schmidt-Lebuhn AN, Knerr NJ, Kessler M. Non-geographic collecting biases in herbarium specimens of Australian daisies (Asteraceae). Biodivers Conserv. 2013; 22: 905–19.
- Yang W, Ma K, Kreft H. Environmental and socio-economic factors shaping the geography of floristic collections in China. Glob Ecol Biogeogr. 2014; 23: 1284–92.
- 37. Efron B. Bootstrap methods: another look at the jackknife. Ann Stat. 1979; 7: 1–26.
- 38. Smith EP, van Belle G. Nonparametric estimation of species richness. Biometrics. 1984; 40: 119–29.
- **39.** Smyth A, Friedel M, O'Malley C. The influence of buffel grass (Cenchrus ciliaris) on biodiversity in an arid Australian landscape. Rangel J. 2009; 31: 307.
- Miller G, Friedel M, Adam P, Chewings V. Ecological impacts of buffel grass (Cenchrus ciliaris L.) invasion in central Australia—does field evidence support a fire-invasion feedback? Rangel J. 2010; 32: 353.
- 41. Guerin GR, Lowe AJ. "Sum of inverse range-sizes" (SIR), a biodiversity metric with many names and interpretations. Biodivers Conserv. 2015; 24: 2877–82.

- **42.** Webb CO, Donoghue MJ. Phylomatic: tree assembly for applied phylogenetics. Mol Ecol Notes. 2005; 5: 181–3.
- **43.** Wikström N, Savolainen V, Chase MW. Evolution of the angiosperms: calibrating the family tree. Proc R Soc London B. 2001; 268: 2211–20.
- Webb CO. Exploring the phylogenetic structure of ecological communities: an example for rain forest trees. Am Nat. 2000; 156: 145–55. PMID: 10856198
- 45. Faith DP. Conservation evaluation and phylogenetic diversity. Biol Cons. 1992; 61: 1–10.
- 46. Rosauer D, Laffan SW, Crisp MD, Donnellan SC, Cook LG. Phylogenetic endemism: A new approach for identifying geographical concentrations of evolutionary history. Mol Ecol. 2009; 18: 4061–72. doi: 10.1111/j.1365-294X.2009.04311.x PMID: 19754516
- Guerin GR, Lowe AJ. Mapping phylogenetic endemism in R using georeferenced branch extents. SoftwareX. 2015, in press; doi: <u>10.1016/j.softx.2015.10.002</u>
- Guerin GR, Biffin E, Lowe AJ. Spatial modelling of species turnover identifies climate ecotones, climate change tipping points and vulnerable taxonomic groups. Ecography. 2013; 36: 1086–96.
- Palmer MW. Putting things in even better order: the advantages of Canonical Correspondence Analysis. Ecology. 1993; 74: 2215–2230.
- 50. Legendre P. Interpreting the replacement and richness difference components of beta diversity. Glob Ecol Biogeogr. 2014; 23: 1324–34.
- 51. R Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing; 2014.
- 52. AW NRM Board. Alinytjara Wilurara Regional NRM Plan. Adelaide; 2010.
- 53. Read JL, Ward MJ. Bringing back warru: initiation and implementation of the South Australian Warru Recovery Plan. Aust Mammal. 2011; 33: 214.
- Andersen AN, Del Toro I, Parr CL. Savanna ant species richness is maintained along a bioclimatic gradient of increasing latitude and decreasing rainfall in northern Australia. J Biogeogr. 2015; 42: 2313– 2322.
- Buffel Grass and fire in a Gidgee and Brigalow woodland: A case study from central Queensland. Ecol Manag Restor. 2003; 4: 120–5.
- **56.** Clarke PJ, Latz PK, Albrecht DE. Long-term changes in semi-arid vegetation: Invasion of an exotic perennial grass has larger effects than rainfall variability. J Veg Sci. 2005; 16: 237–48.